Oxidative desulfurization—fluorination of thioethers. Application for the synthesis of fluorinated nitrogen containing building blocks†‡

Verena Hugenberg, Roland Fröhlich and Günter Haufe*

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An oxidative desulfurization—fluorination protocol has been used to synthesize (2*S*)-2-(difluoromethyl)-*N*-tosylpyrrolidine (**6a**) and (2*S*)-2-(trifluoromethyl)-*N*-tosylpyrrolidine (**7a**) from the (2*S*)-prolinolderived (2*S*)-2-(4-chlorophenylthiomethyl)-*N*-tosylpyrrolidine (**9**) or (2*S*)-2-(dithian-2-yl)-*N*-tosylpyrrolidine (**5**). Efforts to prepare 3,3-difluoroalanine similarly from an *N*-protected *S*-aryl-cysteine ester **17** gave only traces of the target compound **18**. Instead, an unique N-(α , α -difluorobenzyl)-N- α ', α '-dibromoglycine ester **19** was formed by an unprecedented sequence of reaction steps. A plausible mechanism is suggested involving a sulfur-assisted deoxygenation-difluorination of an imino oxygen and a haloform reaction like carbon—carbon bond fission as key-steps. Efforts to prepare (2*S*)-2-(fluoromethyl)-*N*-tosylpyrrolidine (**12**) from (2*S*)-*N*-tosylprolinol (**3**) by treatment with FluoleadTM (1-tert-butyl-4-trifluorosulfanyl-3,5-dimethylbenzene) gave only 5% of the target compound, but 95% of (3*R*)-3-fluoro-*N*-tosylpiperidine (**11a**) by ring enlargement.

Introduction

Among others, amino acids, peptides and amines play important roles in biological processes. Since a couple of years medicinal chemists use specific fluorine substitution to modify the bioavailability and the metabolism of medicinally relevant compounds, but also the affinity and selectivity of the interaction of small fluorinated molecules with biomacromolecules.^{2,3}

The electron-withdrawing effect of a fluorine atom or a fluoroalkyl group modifies the pK_a of neighboring functions, and hence their character as hydrogen bond donors or acceptors. At lower p K_a protonation of amino functions becomes more difficult. The decreased basicity may alter the receptor affinity depending on whether the ligand acts in its neutral or in the protonated form. By way of example, MAO inhibitors were developed by applying the p K_a lowering effect of fluorine to amines, e.g. (E)-2-(3,4-dimethoxyphenyl)-3-fluoroallylamine was found to be an irreversible inhibitor of MAO with good MAO B selectivity.⁴ The consequences of fluorine substitution on the MAO affinity and selectivity of fluorinated tranyleypromine derivatives have been extensively studied by our group.⁵ Also the resorption properties of a molecule can be modified by the influence of fluorine atoms on the p K_a of neighboring ionizable functions and by lipophilicity effects. The uptake process of an ionizable drug depends on the respective proportions and lipophilicity of charged and neutral species. The introduction of fluorine atoms may allow modulation of the ionization of a molecule at physiological pH of 7.4. Thus, lowering of the pK_a of amines and nitrogen-containing heterocycles, by means of fluorinated substituents, can be a very important factor to facilitate the bioavailability, especially for oral administration of drugs.⁶

Therefore, the synthesis of fluorinated amines and amino acids is of particular interest.7 Also different methods for the preparation of α,α-difluoro amino acids are known. The majority of these synthetic strategies apply building blocks for the introduction of difluoromethyl or difluoromethylene moieties. The most frequently used building block is the Reformatsky analogous reagent BrZnCF2CO2Et, which adds to aldehydes, 8-10 imines, 11,12 oxazolidines¹³ or sulfinimines.^{14,15} 2,2-Difluoromethylornithine (elflornithine) was synthesized from CHClF2 and the resulting difluoro derivatives were then transformed to the gem-difluorinated amino acid.16 In order to obtain L-4,4-difluoroglutamic acid gem-difluorinated precursors were synthesized by electrophilic fluorination with N-fluorobenzenesulfonimide (NSFI)¹⁷ or by fluorination of (R)-2,3-O-isopropylidene glyceraldehydes with morpholino trifluorosulfurane (Morpho-DAST).18 Attempts of direct nucleophilic fluorination of keto esters19 with DAST only led to 20-31% of the difluorinated amino acids. By fluorodesulfurization reaction of cysteine with elemental fluorine, 33% of a mixture of 3-fluoro- and 3,3-difluoroalanine (92:8) was obtained. 20 Using trifluoromethyl fluoroxytrifluoromethane or perchlorylfluoride α,α -diffuorinated β -amino acids were formed. With SF₄ and DAST mono- and difluorination of protected amino acids with hydroxyl and oxo functions are possible. 23,24

However, the described fluorination agents for the direct fluorination of amino acids are difficult to handle or did not lead selectively to the geminal difluorinated products. In this paper we report about the oxidative desulfurization—difluorination as a method for the preparation of α,α -difluoromethyl substituted amines and amino acids.

Results and Discussion

Fluoroalkylated pyrrolidine derivatives have been shown to be interesting for the development of new caspase inhibitors.^{25,26}

Organisch-Chemisches Institut and European Institute for Molecular Imaging, Westfälische Wilhelms-Universität, Münster, Germany. E-mail: haufe@uni-muenster.de; Fax: 49 251 83 39772; Tel: 49 251 83 33281

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(2S)-2-(1,3-dithian-2-yl)-N-tosyl-pyrrolidine (5) seemed to be a suitable precursor for initial experiments to synthesize difluoromethyl substituted pyrrolidine derivatives by desulfurization fluorination. Several protocols for similar reactions with different substrates have been published already²⁷ including our own results.²⁸ In a five step reaction sequence 5 was prepared from Lproline (1), which initially was N-protected with 4-toluenesulfonyl chloride and sodium bicarbonate. The formed N-tosyl-L-proline (2) was reduced to N-tosyl-prolinol (3) with sodium borohydride in the presence of borontrifluoride diethyl etherate. The alcohol 3 was converted to the aldehyde 4 by Swern-oxidation. Subsequent Corey-Seebach reaction with 1,3-propanedithiol led to the dithiane 5 with an overall yield of 27% (Scheme 1).

Scheme 1 Synthesis of (2S)-2-(1,3-dithian-2-yl)-N-tosylpyrrolidine (5).

In order to synthesize the difluoromethyl substituted pyrrolidine 6a, the dithiane 5 was reacted with DBH (3.0 equivalents) and Olah's reagent (2.2 equivalents) in dry dichloromethane. After work up and column chromatography the gem-difluoride 6a was obtained as the major product (19 F NMR). The trifluoride 7a and the N-deprotected difluorinated and trifluorinated proline derivatives **6b** and **7b** were detected as by-products (Table 1, entry 1). After treatment of the whole product mixture of entry 1 with tosylchoride in the presence of a base (entry 1a), (2S)-2-difluoromethyl-N-tosylpyrrolidine (6a) was obtained as the main product (90%, ¹⁹F NMR) and (2S)-2-trifluoromethyl-Ntosylpyrrolidine (7a) as the only by-product (10%, ¹⁹F NMR). The product mixture was isolated as a colorless waxy solid (61% yield of 6a). Due to similar R_i -values the separation of the two substances by column chromatography was not possible.

Variation of the reaction conditions to 30 min at 0 °C and neutralization of the reaction mixture by column filtration through basic alumina led to a higher ratio of the trifluoride 7a in the product mixture (entry 2). Again complete separation of the two compounds 6a and 7a was not possible, but the trifluoride 7a could be obtained in an enriched mixture (7a:6a = 65:35, yield: 20%). After further optimization towards compound 7a, may be by enhanced excess of Olah's reagent, this method is applicable for the preparation of compounds with potential biological relevance. The only yet published synthesis of (2S)-2-trifluoromethyl-Ntosylpyrrolidine (7a) was reported by Shustov et al.²⁹ The authors used a fluorodesoxygenation reaction of (S)-proline with SF₄ in HF giving 28% of 7a. For this reaction a steel autoclave, as well as special equipment for the handling with SF₄ and HF are necessary. A further disadvantage of this reaction was the long reaction time of 8 h. In contrast our synthesis can be performed in simple PTFE-flasks and shows a complete conversion of 5 within 30 min at 0 °C.

Good selectivity for **6a** was observed at room temperature (entries 3 and 4) but at longer reaction time 29% of several not identified fluorinated by-products were formed (19 F NMR). Structural assignment of (2S)-2-difluoromethyl-N-tosylpyrrolidine (6a) was difficult on a first view because of the different coupling patterns of the two diastereotopic fluorine atoms F1 and F2 (see Fig. 1). Fluorine atom F1 has a chemical shift of $\delta = -122.6$ ppm and splits to a four-line AB-spectrum, without a vicinal F,H-coupling, while F2 forms an AB-spectrum with eight lines at $\delta = -136.8$ ppm induced by an additional $^{3}J_{\rm EH} = 25.3$ Hz by coupling with the proton of the CH-group of the proline ring. The arrangement of the difluoromethyl moiety can also be confirmed by the coupling of the CH-proton of the proline ring in the ¹H NMR spectrum. This proton shows a coupling pattern of a dublett of a multiplett with only one H,F-coupling of ${}^{3}J_{H,F} = 25.6$ Hz at $\delta =$ 3.79 ppm.

The different coupling pattern of the fluorine atoms is probably caused by the preferred conformation of the compound shown in Fig. 1. The difluoromethyl group is arranged in such a way that the dihedral angel between F1 and the proton of the CHgroup of the proline ring is perpenticular and consequently no coupling of these two nuclei can be observed. The second fluorine atom F2 is located in a gauche position to the proline proton and shows a coupling constant of ${}^{3}J_{\rm F,H} = 25.3$ Hz. The structure shown in Fig. 1 was geometry optimized by quantum chemical calculation (B3LYP/6-311+G(2d,2p)). The calculation did not exhibit an exact orthogonal geometry of one fluorine atom to the β-proton.

 Table 1
 Oxidative desulfurization-diffuorination reactions of dithianen 5

$$\begin{array}{c} 3.0 \text{ eq. DBH} \\ 2.2 \text{ eq. Py. 9HF} \\ \hline \\ 5 \end{array}$$

		Py-9HF (eq.)	Reaction conditions	Crude Product Mixture (19 F NMR,%)				
Entry	DBH (eq.)			6a	6b	7a	7b	others
1"	3.0	2.2	-78 °C; 30 min, -60 °C; 1 h, 0 °C; 30 min, r.t.	59	30	8	3	_
2	3.0	2.2	30 min, 0 °C (Alumina)	56	_	44	_	_
3	3.0	2.2	30 min, r.t. (Alumina)	95 ^b	_	5 ^b	_	_
4	3.0	2.2	20 h, r.t.	65	_	6	_	29

^a After tosylation of the whole product mixture of entry 1 a 90:10 mixture (19F NMR) of 6a and 7a was found. ^b After column chromatography.

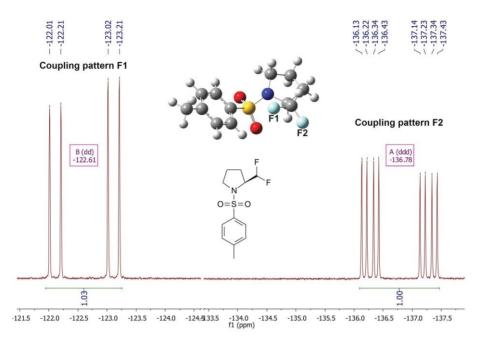


Fig. 1 Coupled ¹⁹F NMR spectra of (2S)-2-difluoromethyl-N-tosylpyrrolidine (6a).

The formation of the *gem*-diffuoro compound **6a** can be formulated analogously to the fluorodesulfurization mechanism of dithianes postulated by Katzenellenbogen *et al.*^{27b} The probable mechanism of the formation of the trifluoride **7a** is analogous to the one postulated in our previous work.²⁸

In order to avoid the five-step preparation of (2S)-N-tosyl-2-(dithian-2-yl)pyrrolidine (5), we intended to apply an oxidative desulfurization—difluorination approach^{28,30} for the synthesis of (2S)-2-difluoromethyl-N-tosylpyrrolidine (6a). Therefore, (2S)-(4-chlorophenylthiomethyl)-N-tosylpyrrolidine (9) was prepared from N-tosyl-L-prolinol (3) by O-tosylation to form 8 and subsequent nucleophilic substitution of the tosylate with p-chlorothiophenol (Scheme 2). The formed thioether 9 was then used as a starting material for fluorination.

Scheme 2 Synthesis of (2S)-(4-chlorophenylthiomethyl)-N-tosylpyrrolidine (9).

Applying our standard oxidative desulfurization—difluorination conditions,²⁸ **9** was reacted with 3 equivalents of DBH and 6 equivalents of Py·9HF in dry CH₂Cl₂ at room temperature for 17 h (Table 2, entry 1). Filtration of the product mixture over basic alumina and column chromatography (silica gel) afforded (2S)-2-difluoromethyl-N-tosylpyrrolidine (**6a**) as a waxy colorless substance with 33% yield. Additionally, (2S)-2-dibromofluoromethyl-N-tosylpyrrolidine (**10**) (13%) and the 3-fluoro-piperidine derivative **11a** (16%), were isolated. Lowering the amount of both the electrophile and the fluorinating reagent to 2 equivalents of DBH and 3 equivalents of Olah's reagent caused the preferred formation of the *gem*-dibromofluoride **10**. The *gem*-difluoride **6a** (15%) was detected as a by-product (entry

2). With 2 equivalents of DBH and 4 equivalents of Py·9HF the *gem*-dibromofluoride **10** was formed almost exclusively and (2S)-2-fluoromethyl-N-tosylpyrrolidine (**12**) was detected as the only by-product (entry 3, for the mechanism of formation see ref. 28b).

In contrast, increasing the amount of the fluorinating reagent to 9 equivalents in combination with 3 equivalents of DBH led preferably to the fluorinated piperidine derivative 11b, which is *meta*-brominated at the tosyl ring. In addition, a small amount of dibromofluoride 10 was identified.

Compounds **6a**, **7a**, and **10** are formed by initial fluoro-Pummerer reaction(s) and subsequent desulfurization—fluorination or —bromination as discussed earlier for similar transformations of a variety of alkyl aryl thioethers.²⁸

(3R)-3-Fluoro-N-tosylpiperidine (11a) itself could be synthesized selectively from N-tosyl-L-prolinol (3) using the new fluorinating reagent³¹ FluoleadTM (2 equivalents) and Olah's reagent (0.22 equivalents) with an isolated yield of 95% (Scheme 3).

Scheme 3 Mechanism of the conversion of 3 with Fluolead $^{\text{TM}}$ and Olah's reagent.

The formation of 11a proceeds via a ring expansion reaction similar to the one proposed by Shreeve et al. 32 and Cossy

Table 2 Oxidative desulfurization–diffuorination of (2S)-2-(4-chlorophenylthiomethyl)-N-tosylpyrrolidine (9)

			Reaction Conditions	Crude P	roduct Mixture			
Entry	DBH (eq.)	Py-9HF (eq.)		6a	7a	10	11a/11b	12
1	3.0	6.0	17 h, r.t.	54	2	14	31"	_
2	2.0	3.0	17 h, r.t.	15	_	85	_	_
3	2.0	4.0	17 h, r.t.	_	_	90	_	10
4	3.0	9.0	17 h, r.t.	1	_	7	92 ^b	_

^a 3-fluoro-*N*-tosylpiperidine (11a). ^b 3-fluoro-*N*-(3-bromotoluenesulfonyl)piperidine (11b)

et al.33 for reactions of prolinol derivatives, which with DAST or desoxofluorTM lead to mixtures of optical active fluoro pyrrolidine and fluoro piperidine derivatives. The selectivity of these rearrangements depended on the substituents at the pyrrolidine ring and steric constrains on nitrogen. Using FluoleadTM as a fluorinating agent we detected only 5% of the not rearranged product (2S)-2-fluoromethyl-N-tosylpyrrolidine (12) in the product mixture (¹⁹F NMR). Thus, the selectivity and the yield (95% of 11a) of this reaction are much better than most of the reactions described in literature. 32,33

The oxidative desulfurization-difluorination might also be an opportunity for the direct difluorination of amino acids in β-position starting from suitably substituted amino acids such as S-phenylcysteine derivatives. In order to prepare 3,3difluoroalanine on an alternative pathway to the use of fluorinated building blocks,34-37 or fluorinating reagents like fluoroxytrifluoromethane or elemental fluorine²⁰ we synthesized methyl (2S)-2-(1,3-dioxoisoindolin-2-yl)-3-(phenylthio)propanoate (17) as a precursor for an oxidative desulfurization-difluorination reaction in a three step protocol starting from L-serine. After protection of the amino function with N-ethoxycarbonylphthalimide and Na₂CO₃ to form 14 the carbonyl function was esterified with methanol to yield 15. The hydroxyl function was scheduled to be converted to a tosyl group. Unfortunally, elimination of toluenesulfonic acid to 16³⁸ took place during purification on silica gel or basic alumina due to the high acidity of the proton in α -position of the nitrogen. Finally, the reaction of 15 with Hata's reagent (diphenyldisulfide and tributylphosphine)39 afforded the target compound, methyl (2S)-2-(1,3-dioxoisoindolin-2-yl)-3-(phenylthio)propanoate (17), in 69% yield (Scheme 4).

In the next reaction step, 17 was subjected to the standard conditions of the oxidative desulfurization–difluorination (3 equivalents DBH, 6 equivalents Py-9HF). 28 However, the expected difluorinated product, methyl (2S)-2-(1,3-dioxoisoindolin-2-yl)-3,3-difluoropropanoate (18), was only verified as a by-product by ¹⁹F NMR spectroscopy and ESI mass spectrometry (Scheme 5). The major product of the reaction was isolated by column chromatography on neutral alumina. Initially, no structure could be proposed due to the lack of indicative signals in ¹H, ¹³C, and ¹⁹F NMR spectra for structure elucidation. Finally, repeated recrystallization from pentane-diethyl ether led to crystals suitable for X-ray crystallographic analysis. According to these data, the prod-

Scheme 4 Synthesis of methyl (2S)-2-(1,3-dioxoisoindolin-2-yl)-3-(phenylthio)propanoate (17).

Scheme 5 Oxidative desulfurization—diffuorination reaction of 17.

uct was identified as the α , α -dibromo- α' , α' -difluoroalkylamide 19 (Fig. 2). The amide nitrogen is flanked by a difluoromethylene group on the one and a dibromomethylene moiety on the other side. This structure unit to the best of our knowledge was not known in literature before.

This particular product necessitated some considerations about its formation from 17. The formal exchange of a carboxyl oxygen atom as part of an imino group by two fluorine atoms with DBH and Olah's reagent was not observed till now. Generally fluorinating agents like SF₄, DAST or desoxofluorTM are necessary for the direct transformation of carbonyl oxygen atoms to gem difluorides. 40,41 Also the introduction of two bromine atoms in α position of the nitrogen of an amino acid formally replacing an arylthiomethyl moiety was not described yet.

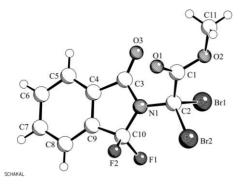


Fig. 2 X-ray structure of methyl 2,2-dibromo-2-(1,1-difluoro-3oxoisoindolin-2-yl)acetate (19).

Thus, we speculated about a plausible mechanism of formation of the α,α -dibromo- α',α' -difluoroamide 19 (Scheme 6). Initially, the sulfur is attacked by the electrophile DBH (Br+) to form intermediate I. Due to the close proximity of the sulfenium ion center to one of the phthalimide oxygens, nucleophilic attack of the oxygen on sulfur can occur, analogously to the reaction of a ketone with DAST, whereupon via an oxonium ion the stabilized carbenium ion II can be formed. The carbenium ion II can add a fluoride from the fluorinating reagent forming a carbon flanked by fluorine, oxygen and an imide nitrogen. This strongly electrophilic carbon is attacked again by a fluoride. By breaking the carbon-oxygen bond, bromide is eliminated and the sulfoxide, methyl (2S)-2-(1,1-difluoro-3-oxoisoindolin-2-yl)-3-(phenylsulfinyl)propanoate (20) is formed.

The two bromine atoms might be introduced subsequently on the following way: A second bromonium ion attacks sulfur to form the cation III. HBr elimination from III leads to the carbenium/sulfoxonium ion IV, from which the olefin 21 is formed by deprotonation. Addition of Br₂ or "BrF" to the double bond of 21, elimination of hydrogen bromide and again addition of "BrX" to the double bond leads to the trihalogen moiety of 22. In the course of a "haloform" reaction a dihalogenmethyl sulfinyl anion is eliminated simultaneously with the attack of a bromide on 22 to obtain the α,α -dibromo- α',α' -difluoroalkylamide 19. Alternative mechanisms are possible, but seem less probable.⁴²

Conclusion

Up to present there were no simple methods known for the synthesis of (2S)-2-difluoromethylpyrrolidine derivatives. Starting from (2S)-2-(dithian-2-yl)-N-tosylpyrrolidine (5) the selective synthesis of (2S)-2-difluoromethyl-N-tosylpyrrolidine (6a) (61% yield), besides a minor amount of (2S)-2-trifluoromethyl-Ntosylpyrrolidine (7a), was successful by conversion with DBH and Py-9HF. The product ratio was not significantly changed by modified reaction conditions. The share of (2S)-2-trifluoromethyl-N-tosylpyrrolidine (7a) could be slightly increased under the conditions shown in Table 1, entry 2. Complete separation of the two fluorinated pyrrolidine derivatives was not possible.

(2S)-2-Difluoromethyl-N-tosylpyrrolidine (6a) was also synthesized by the oxidative desulfurization-difluorination approach starting from 2-(arylthiomethyl)pyrrolidine 9, however with only 33% yield. Despite the low yield this protocol provides a similar overall yield like the multi-step method via the dithiane 5 (17% or 19%, respectively).

Lowering the amounts of electrophile and fluorinating reagent led exclusively to the dibromofluoride 10. Increasing the amount of Olah's reagent implicated the formation of the ring expanded monofluoro substituted piperidine 11b, monobrominated in metaposition at the tosyl group. The reaction of N-tosyl-L-prolinol (3) with FluoleadTM and Olah's reagent delivered the (3R)-3-fluoro-*N*-tosylpiperidine (11a) in 95% yield by ring enlargement.

Under the conditions of the desulfurization-difluorination reaction with DBH and Olah's reagent of methyl (2S)-2-(1,3dioxoisoindolin-2-yl)-3-(phenylthio)propanoate (17) the expected 3,3-difluoroalanine derivative 18 was formed only as a by-product. The reaction led mainly to a unique, so far not known structure motiv of an α,α -dibromo- α',α' -difluoroalkylamide 19. The

Plausible mechanism of formation of the α,α -dibromo- α',α' -difluoroamide 19.

formation of this product seems to be caused by the close proximity of one of the carboxyl oxygens of the phthalimide moiety to the sulfur initiating a fluoro-Pummerer-like rearrangement.

Experimental

General methods

Column chromatography was performed on Merck silica gel 60 (0.040-0.063 mm). NMR spectra were recorded on a Bruker ARX300 and a Bruker DPX300 (1H NMR, 300 MHz, 13C NMR, 75 MHz, ¹⁹F NMR, 282 MHz), Bruker AMX 400 (¹H NMR, 400 MHz, ¹³C NMR, 100 MHz) and Varian Inova (¹H NMR, 500 MHz, ¹³C NMR, 126 MHz, ¹⁹F NMR, 470 MHz) spectrometers. TMS (1H), CDCl₃ (13C) and CFCl₃ (19F) were used as internal standards. Mass spectra were recorded on Thermo-Finningan MAT8200 (EI, 70 eV), Waters-Micromass GCT (GCToF, EI), and Waters-Micromass Quatromicro GC (GC/CI and EI, 70 eV) instruments. All air and moisture-sensitive reactions were performed under argon atmosphere. Solvents were purified and dried by literature methods where necessary. The reactions with Olah's reagent were performed in TeflonTM flasks. The alkyl aryl thioethers were prepared from the corresponding thiophenols and alkyl halides under basic conditions.⁴³

Synthesis of (2S)-2-(dithian-2-yl)-N-tosylpyrrolidine (5)

N-Tosyl-L-proline (2) (100%) and *N-tosyl-L-prolinol* (3) (89%) were prepared according to literature procedures.⁴⁴ *N-Tosyl-L-prolinal* (4) was synthesized by Swern-oxidation in a 23.0 mmol scale (5.83 g, 100%) and isolated as a yellow solid; mp 117 °C with decomposition (lit.,⁴⁵ 139-141 °C); α _D²⁰ -102.0 (α 1.03 in CHCl₃) (lit.,⁴⁵ α _D²⁰ -121.0 (α 1.00 in MeOH). The spectroscopic data match with those given in literature.⁴⁵

(2S)-2-(dithian-2-yl)-N-tosylpyrrolidine (5) was synthesized by Corey-Seebach reaction according to the literature procedure in a 5.87 mmol scale.46 The product was purified by column chromatography (silica gel, cyclohexane/ethyl acetate, 4:1) and obtained as a yellowish solid (624 mg, 31%); mp 97 °C; $[\alpha]_D^{20}$ -72.3 (c 0.98 in CHCl₃); $\delta_{\rm H}$ (300 MHz; CDCl₃, TMS): 1.44 (1 H, m, CH₂), 1.69 (1 H, m, CHCH₂), 1.76–1.94 (2 H, m, CH₂), 2.07–2.18 (2 H, m, CH₂), 2.43 (3 H, s, CH₃), 2.82–3.00 (4 H, m, SCH₂), 3.20 (1 H, m, NCH₂), 3.39 (1 H, m, NCH₂), 3.88 (1 H, m, NCH), 4.74 (1 H, d, ${}^{3}J_{H,H}$ = 4.0 Hz, SCHS), 7.32 (2 H, d, ${}^{3}J_{H,H}$ = 8.0 Hz, Ph-CH), 7.76 (2 H, d, ${}^{3}J_{H,H}$ = 8.3 Hz, Ph-CH); δ_{C} (75 MHz; CDCl₃): 21.5 (CH₃), 24.9 (CH₂), 26.2 (CH₂), 28.2 (CHCH₂), 30.1 (SCH₂), 30.9 (SCH₂), 49.8 (NCH₂), 54.3 (SCHS), 62.6 (NCH), 127.6 (Ph-CH), 129.7 (Ph-CH), 134.4 (Ph-C), 143.5 (Ph-C). MS (EI-GCinlet): m/z (%) 343 (<0.1) [M⁺], 224 (100) [C₁₁H₁₄NO₂S⁺], 155 (24) $[C_7H_7O_2S^+]$, 119 (10) $[C_4H_7S_2^+]$, 91 (67) $[C_7H_7^+]$. Exact mass (ESI): [M+Na⁺] calcd for C₁₈H₂₆FNO₅SNa⁺: 366.0632; found: 366.0625. Anal. calcd. for C₁₅H₂₁NO₂S₃: C, 52.44; H, 6.16; N, 4.08. Found: C, 52.11; H, 5.94; N, 3.99.

Conversion of (2S)-2-(dithian-2-yl)-N-tosylpyrrolidine (5) with DBH and Olah's reagent

Synthesis of (2*S*)-2-difluoromethyl-*N*-tosylpyrrolidine (6a) (Table 2, entry 1). 5,5-Dimethyl-1,3-dibromohydantoin (DBH, 172 mg, 0.60 mmol, 3.0 eq.) was dissolved in dry dichloromethane

(4 cm³) under argon in a flame dried PTFE-flask and cooled to -78 °C. Then Olah's reagent (0.10 cm³, 0.44 mmol, 2.2 eq.) was dropped slowly to the reaction mixture. Subsequently 5 (69 mg, 0.20 mmol), dissolved in dry dichloromethane (2 cm³), was dropped to the reaction mixture within 10–15 min. The mixture was stirred for 30 min at -60 °C, 1 h at 0 °C and 30 min at room temperature. Afterwards it was cooled down to 0 °C and neutralized with ice-cold saturated aqueous NaHCO₃ solution. The aqueous phase was extracted with dichloromethane $(3 \times 4 \text{ cm}^3)$ and the combined organic layer was washed with 1 N HCl, 5% aqueous NaHCO₃ and brine $(2 \times 5 \text{ cm}^3)$ and dried over anhydrous MgSO₄. After concentration under reduced pressure, the products were separated by column chromatography (silica gel, pentane-diethyl ether, 10:1). The main product of this reaction was the tosyl protected (2S)-2-difluoromethylpyrrolidine derivative 6a. As byproducts the deprotected (2S)-2-difluoromethylpyrrolidine (6b) and the N-protected and the deprotected (2S)-2-trifluoromethyl pyrrolidine derivatives 7a and 7b were found.

In a second attempt, the whole product mixture was subjected to tosylation according to the method given in ref. ⁴⁴ (Table 1, entry 1a). Under these conditions **6a** was the main product (90%, ¹⁹F NMR) and **7a** (10%, ¹⁹F NMR) was the only by-product. Column chromatographic separation of the by-product was not possible. The product mixture was obtained as a colorless wax (yield calculated for **6a**: 34 mg, 61%).

(2S)-2-difluoromethyl-N-tosylpyrrolidine (6a). $[\alpha]_D^{20}$ -38.7 (c 0.70 in CHCl₃); $\delta_{\rm H}$ (500 MHz, CDCl₃ TMS): 1.50–1.67 (2 H, m, CH₂ & CHCH₂), 1.92 (1 H, m, CH₂), 2.10 (1 H, m, CHCH₂), 2.45 (3 H, s, CH₃), 3.15 (1 H, m, NCH₂), 3.47 (1 H, m, NCH₂), 3.79 (1 H, dm, ${}^{3}J_{HF} = 25.6$ Hz, NCH), 6.11 (1 H, ddd, ${}^{2}J_{HF} = 57.9$ Hz, ${}^{2}J_{HF} =$ 55.0 Hz, ${}^{3}J_{H,H} = 1.6$ Hz, CF₂H), 7.35 (2 H, d, ${}^{3}J_{H,H} = 8.0$ Hz, Ph-CH), 7.73 (2 H, m, ${}^{3}J_{HH} = 8.3$ Hz, Ph-CH); δ_{C} (126 MHz, CDCl₃): $21.6 (CH_3), 24.3 (d, {}^{3}J_{C,F} = 4.3 Hz, CHCH_2), 24.6 (d, {}^{4}J_{C,F} = 2.1 Hz,$ CH₂), 49.3 (NCH₂), 60.0 (dd, ${}^{2}J_{CF} = 31.8$ Hz, ${}^{2}J_{CF} = 22.5$ Hz, NCH), 115.8 (dd, ${}^{1}J_{C,F} = 247.6$ Hz, ${}^{1}J_{C,F} = 241.7$ Hz, CF₂H), 127.6 (Ph-CH), 129.9 (Ph-CH), 133.7 (Ph-C), 144.1 (Ph-C); $\delta_{\rm F}$ $(282 \text{ MHz}, \text{CDCl}_3, \text{CFCl}_3): -122.6 (1 \text{ F}, \text{dd}, {}^2J_{\text{EF}} = 283.5 \text{ Hz}, {}^2J_{\text{HF}} =$ 54.9 Hz), -136.9 (1 F, ddd, ${}^{2}J_{E,F} = 283.4$ Hz, ${}^{2}J_{H,F} = 58.0$ Hz, $^{3}J_{HF} = 25.3 \text{ Hz}$). MS (EI-GC-inlet): m/z (%) 275 (<0.1) [M⁺], 224 (59) $[C_{11}H_{14}NO_2S^+]$, 155 (40) $[C_7H_7O_2S^+]$, 91 (100) $[C_7H_7^+]$, 65 (44), 41 (19) $[C_3H_5^+]$. Exact mass (ESI): $[M+Na^+]$ calcd for $C_{12}H_{15}F_2NO_2SNa^+$: 298.0684; found: 298.0680.

Synthesis of (2S)-2-trifluoromethyl-N-tosylpyrrolidine (7a) (Table 2, entry 2). DBH (412 mg, 1.44 mmol, 3.0 eq.) was dissolved in dry dichloromethane (3 cm³) under argon in a flame dried PTFE-flask and cooled to 0 °C. Olah's reagent (0.24 cm³, 1.06 mmol, 2.2 eq.) and 5 (165 mg, 0.48 mmol), dissolved in dry dichloromethane, were given dropwise to the reaction mixture. The mixture was stirred 30 min at 0 °C and dichloromethane (20 cm³) was added. Then the entire mixture was given to a PE-column filled with basic alumina $(2 \times 10 \text{ cm}, \text{ca } 50 \text{ g})$ for neutralization. The filtrate was collected, the column was rinsed with dichloromethane (20 cm³), the dichloromethane phase was dried over anhydrous MgSO₄ and the solvent was removed under reduced pressure. After column chromatography (silica gel, cyclohexane/ethyl acetate, 20:1) the trifluoride **7a** (65%, ¹⁹F NMR) was obtained in a mixture with the difluoride **6a** as the only by-product (35%, ¹⁹F NMR) as a colorless oil (yield calculated for 7a: 14 mg, 20%).

(2S)-2-trifluoromethyl-N-tosylpyrrolidine (7a). [α]_D²⁰ –38.0 (c 0.67 in CHCl₃); δ_H (500 MHz, CDCl₃, TMS): 1.74 (1 H, m, CHCH₂), 1.86 (1 H, m, CH₂), 1.97 (1 H, m, CHCH₂), 2.07 (1 H, m, CH₂), 2.44 (3 H, s, CH₃), 3.33 (1 H, m, NCH₂), 3.49 (1 H, m, NCH₂), 4.42 (1 H, m, NCH), 7.35 (2 H, d, 3J _{H,F} = 8.0 Hz, Ph-CH), 7.74 (2 H, d, 3J _{H,F} = 8.4 Hz, Ph-CH); δ_C (126 MHz, CDCl₃): 21.5 (CH₃), 26.4 (CH₂), 29.3 (CH₂CH), 49.1 (NCH₂), 59.7 (q, 2J _{C,F} = 32.0 Hz, NCH), 125.2 (q, 1J _{C,F} = 281.6 Hz, CF₃), 127.5 (Ph-CH), 129.8 (Ph-CH), 133.8 (Ph-C), 143.8 (Ph-C); δ_F (282 MHz, CDCl₃, CFCl₃): –75.7 (3 F, d, 3J _{H,F} = 7.6 Hz). MS (EI-GC-inlet): m/z (%) 293 (6) [M⁺], 224 (70) [C₁₁H₁₄NO₂S⁺], 155 (51) [C₇H₇O₂S⁺], 91 (100) [C₇H₇⁺], 65 (37), 41 (19) [C₃H₅⁺]. Exact mass (ESI): [M+Na⁺] calcd for C₁₂H₁₄F₃NO₂SNa⁺: 316.0590; found: 316.0584.

Synthesis of (2S)-2-(4-chlorophenylthiomethyl)-N-tosylpyrrolidine (9)

N,O-Bis-tosyl-L-prolinol (8)⁴⁷. A suspension of N-tosyl-Lprolinol (3) (2.838 g, 11.13 mmol) in dry diethyl ether (16 cm³) was cooled to 0 °C. p-Toluenesulfonyl chloride (2.122 g, 11.13 mmol, 1.0 eq.) and powdered sodium hydroxide (533 mg, 13.35 mmol, 1.2 eq.) were added and the mixture was stirred at 0 °C for 1 h. Stirring was continued over night at room temperature. Then ice water was added (30 cm³) and the aqueous phase was extracted with diethyl ether $(3 \times 20 \text{ cm}^3)$. The combined organic layer was washed with water and brine, dried over MgSO4 and the solvent was removed under reduced pressure. After column chromatography (silica gel, cyclohexane/ethyl acetate, 6:1) the product 8 was obtained as a white solid (3.168 g, 79%); mp 100 °C; $[\alpha]_{D}^{20}$ –119.3 (c 1.00 in CHCl₃); δ_{H} (400 MHz, CDCl₃, TMS): 1.50– 1.70 (2 H, m, CHCH₂), 1.73-1.93 (2 H, m, CH₂), 2.43 (3 H, s, CH₃), 2.47 (3 H, s, CH₃), 3.04 (1 H, m, NCH), 3.39 (2 H, m, NCH_2), 3.75 (1 H, m, NCH_2), 3.95 (1 H, dd, ${}^2J_{H,H} = 9.9 \text{ Hz}, {}^3J_{H,H} =$ 8.2 Hz, CHC H_2 O), 4.25 (1 H, dd, ${}^2J_{H,H}$ = 9.9 Hz, ${}^3J_{H,H}$ = 3.6 Hz, CHC H_2 O), 7.31 (2 H, d, ${}^3J_{H,H}$ = 8.0 Hz, Ph-CH), 7.38 (2 H, d, $^{3}J_{H,H} = 8.0 \text{ Hz}, \text{ Ph-CH}), 7.66 (2 \text{ H}, \text{ d}, ^{3}J_{H,H} = 8.3 \text{ Hz}, \text{ Ph-CH}), 7.82$ $(2 \text{ H}, \text{ d}, {}^{3}J_{\text{H,H}} = 8.3 \text{ Hz}, \text{Ph-CH}); \delta_{\text{C}} (101 \text{ MHz}, \text{CDCl}_{3}): 21.5 (\text{CH}_{3}),$ 21.7 (CH₃), 23.7 (CH₂), 28.5 (CH₂CH), 49.3 (NCH₂), 57.6 (NCH), 71.5 (CHCH₂O), 127.6 (Ph-CH), 128.0 (Ph-CH), 129.8 (Ph-CH), 130.0 (Ph-CH), 132.6 (Ph-C), 133.5 (Ph-C), 143.9 (Ph-C), 145.0 (Ph-C). Exact mass (ESI): $[M+H^+]$ calcd for $C_{19}H_{23}NO_5S_2H^+$: 410.1090; found: 410.1095. Exact mass (ESI): [M+Na+] calcd for C₁₀H₂₃NO₅S₂Na⁺: 432.0910; found: 432.0916. Anal. calcd. for C₁₉H₂₃NO₅S₂: C, 55.72; H, 5.66; N, 3.42. Found: C, 55.61; H, 5.51; N, 3.44.

(2S)-2-(4-Chlorophenylthiomethyl)-N-tosylpyrrolidine (9). According to the literature procedure N,O-bis-tosyl-L-prolinol (8) (1.638 g, 4.00 mmol) was reacted with p-chlorothiophenol (608 mg, 4.22 mmol).⁴³ Then the reaction mixture was diluted with water (300 cm³) and extracted with ethyl acetate (3×50 cm³). The combined organic layer was washed with water, saturated NaHCO₃ solution and brine (2×50 cm³), dried over MgSO₄ and the solvent was removed under reduced pressure. After column chromatography (silica gel, cyclohexane/ethyl acetate, 8:1) the product was obtained as a colorless oil, that crystallized at friction (1.027 g, 67%); mp 67 °C; $[\alpha]_D^{20}$ –311.1 (c 0.98 in CHCl₃); δ_H (400 MHz, CDCl₃, TMS): 1.46–1.70 (2 H, m, CHCH₂), 1.71–1.94 (2 H, m, CH₂), 2.41 (3 H, s, CH₃), 2.77 (1 H, dd, ${}^2J_{H,H}$ =

13.4 Hz, ${}^{3}J_{H,H}$ = 10.4 Hz, CHC H_{2} S), 3.03 (1 H, m, NCH), 3.57

(2 H, m, NCH₂), 3.64 (1 H, m, CHC H_2 S), 7.26 (2 H, d, ${}^3J_{H,H}$ = 8.0 Hz, Ph-CH), 7.32 (2 H, d, ${}^3J_{H,H}$ = 8.9 Hz, Ph-CH), 7.40 (2 H, d, ${}^3J_{H,H}$ = 8.9 Hz, Ph-CH), 7.55 (2 H, d, ${}^3J_{H,H}$ = 8.3 Hz, Ph-CH); $\delta_{\rm C}$ (101 MHz, CDCl₃): 21.5 (CH₃), 23.7 (CH₂), 30.2 (*C*H₂CH), 38.5 (CHCH₂S), 49.7 (NCH₂), 58.7 (NCH), 127.4 (Ph-CH), 129.1 (Ph-CH), 129.7 (Ph-CH), 130.2 (Ph-CH), 131.9 (Ph-C), 133.7 (Ph-C), 133.9 (Ph-C), 143.6 (Ph-C). MS (EI-GC-inlet): m/z (%) 381 (6) [M⁺], 224 (100) [C₁₁H₁₄NO₂S⁺], 155 (71) [C₇H₇O₂S⁺], 91 (38) [C₇H₇⁺], 45 (6) [C₂H₄O⁺]. Exact mass (ESI): [M+H⁺] calcd for C₁₈H₂₀ClNO₂S₂H⁺: 382.0697; found: 382.0706; [M+Na⁺] calcd for C₁₈H₂₀ClNO₂S₂Na⁺: 404.0516; found: 404.0528. Anal. calcd. for C₁₈H₂₀ClNO₂S₂: C, 56.60; H, 5.28; N, 3.67. Found: C, 56.48; H, 4.98; N, 3.67.

Oxidative desulfurization-difluorination of thioethers

General procedure²⁸

Olah's reagent was added to a solution of the corresponding thioether (0.5 mmol) in dry dichloromethane (5 cm³) in a TeflonTM flask via a polypropylene/polyethylene syringe. DBH was added and the mixture was stirred for 17 h at room temperature. For the particular equivalents of Olah's reagent and DBH see Table 2.

Oxidative desulfurization-difluorination of (2S)-2-(4-chlorophenylthiomethyl)-N-tosylpyrrolidine (9). According to the general procedure, (2S)-2-(4-chlorophenylthiomethyl)-1tosylpyrrolidine (9) (191 mg, 0.5 mmol) was reacted with Olah's reagent (0.69 cm³, 3.0 mmol, 6 eq.) and DBH (431 mg, 1.5 mmol, 3 eq.) in dry dichloromethane (5 cm³) for 17 h at room temperature. Then the reaction mixture was passed through a short PE-column with basic alumina for neutralization and the column was rinsed with dichloromethane (100 cm³). The organic layer was collected in two fractions. The first fraction contained a mixture of the difluoromethyl, trifluoromethyl and dibromofluoromethyl substituted pyrrolidine derivatives 6a, 7a and 10 as well as the monofluorinated piperidine 11a. The second fraction contained pure (3R)-fluoro-N-tosylpiperidine (11a). Repeated column chromatography of the first fraction (silica gel, pentane: 1.000cm³; then pentane-diethyl ether, 3:1) delivered the difluormethyl product **6a** and the dibromofluoromethyl derivative **10**.

(2S)-2-Difluoromethyl-N-tosylpyrrolidine (6a). Isolated as a colorless waxy solid (45 mg, 33%); mp 76–77 °C; $[\alpha]_D^{20}$ –40.7 (c 0.85 in CHCl₃). The spectroscopic data are matching to those given above.

(2*S*)-2-Dibromofluoromethyl-*N*-tosylpyrrolidine (10). Isolated as a colorless oil (68 mg, 33%). Also obtained as main product of the reactions of entry 2 (0.25 mmol scale, 56 mg, 54%) and 3 (0.25 mmol scale, 65 mg, 63%) in Table 2; [α]_D²⁰ –112.9 (*c* 0.93 in CHCl₃); $\delta_{\rm H}$ (500 MHz, CDCl₃, TMS): 1.50–1.61 (2 H, m, CH₂), 2.00 (1 H, m, CH₂), 2.12 (1 H, m, CHC*H*₂), 2.23 (1 H, m, CHC*H*₂), 2.45 (3 H, s, CH₃), 3.15 (1 H, m, NCH₂), 3.46 (1 H, m, NCH₂), 4.77 (1 H, ddd, ${}^{3}J_{\rm H,H}$ = 11.0 Hz, ${}^{3}J_{\rm H,H}$ = 8.7 Hz, ${}^{4}J_{\rm H,H}$ = 4.9 Hz, NCH), 7.33 (2 H, d, ${}^{3}J_{\rm H,H}$ = 8.0 Hz, Ph-CH), 7.76 (2 H, d, ${}^{3}J_{\rm H,H}$ = 8.3 Hz, Ph-CH); $\delta_{\rm C}$ (126 MHz, CDCl₃): 21.6 (CH₃), 25.0 (CH₂), 30.3 (d, ${}^{3}J_{\rm C,F}$ = 1.6 Hz, *C*H₂CH), 50.6 (NCH₂), 71.9 (d, ${}^{2}J_{\rm C,F}$ = 17.6 Hz, NCH), 102.1 (d, ${}^{1}J_{\rm C,F}$ = 326.0 Hz, CBr₂F), 127.4 (Ph-CH), 129.7 (Ph-CH), 136.4 (Ph-C), 143.9 (Ph-C); $\delta_{\rm F}$ (282 MHz, CDCl₃). CFCl₃):

-51.3 (1 F, d, ${}^{3}J_{HF} = 10.9$ Hz). Exact mass (ESI): [M+Na⁺] calcd for $C_{12}H_{14}Br_2FNO_2SNa^+$: 439.8947/437.8968/435.8988; found: 439.8947/437.8965/435.8986.

(3R)-3-Fluoro-*N*-tosylpiperidine $(11a)^{32,33}$. Isolated from the above reaction as a white solid (20 mg, 16%); mp 100 °C; $[\alpha]_{p}^{20}$ +12.6 (c 0.96 in CHCl₃); $\delta_{\rm H}$ (400 MHz, CDCl₃, TMS): 1.54–1.69 (2 H, m, CH₂ & CFHCH₂), 1.76 (1 H, m, CFHCH₂), 1.89 (1 H, m, CH₂), 2.43 (3 H, s, CH₃), 2.95 (1 H, m, NCH₂), 3.05–3.10 (2 H, m, NCH₂ & NCH₂CFH), 3.30 (1 H, ddd, ${}^{3}J_{H,F} = 20.1$ Hz, $^{2}J_{H,H}$ = 11.9 Hz, $^{3}J_{H,H}$ = 3.2 Hz, NC H_{2} CFH), 4.67 (1 H, dtt, $^{2}J_{H,F}$ = 47.4 Hz, ${}^{3}J_{H,H} = 6.9$ Hz, ${}^{3}J_{H,H} = 3.4$ Hz, CFH), 7.34 (2 H, d, $^{3}J_{\rm H,H}$ = 8.0 Hz, Ph-CH), 7.65 (2 H, d, $^{3}J_{\rm H,H}$ = 8.3 Hz, Ph-CH); $\delta_{\rm C}$ (101 MHz, CDCl₃): 20.9 (d, ${}^{4}J_{CF} = 6.4$ Hz, CH₂CH₂CH₂), 21.4 (CH_3) , 29.1 (d, ${}^3J_{CF} = 20.1 \text{ Hz}$, CH_2CFH), 45.6 (NCH₂), 49.5 (d, $^{2}J_{CF} = 26.4 \text{ Hz}, \text{ N}CH_{2}\text{CFH}), 85.9 \text{ (d, }^{1}J_{CF} = 176.4 \text{ Hz}, \text{ CFH}),$ 127.4 (Ph-CH), 129.6 (Ph-CH), 133.1 (Ph-C), 143.6 (Ph-C); $\delta_{\rm F}$ (282 MHz, CDCl₃, CFCl₃): -183.3 (1 F, m). MS (EI-GC-inlet): m/z (%) 257 (38) [M⁺], 237 (9) [M⁺-HF], 224 (2) [C₁₁H₁₄NO₂S⁺], 155 (38) $[C_7H_7O_2S^+]$, 102 (100) $[C_5H_9NF^+]$, 91 (50) $[C_7H_7^+]$. Exact mass (ESI): [M+H⁺] calcd for C₁₂H₁₆FNO₂SH⁺: 258.0959; found: 258.0959. Exact mass (ESI): [M+Na⁺] calcd for C₁₂H₁₆FNO₂SNa⁺: 280.0778; found: 280.0774.

(3R)-N-(3-Bromo-4-toluenesulfonyl)-3-fluoropiperidine Isolated from the reaction shown in Table 2, entry 4 (0.25 mmol scale; 43 mg, 51%); $[\alpha]_D^{20}$ +13.4 (c 1.05 in CHCl₃); δ (400 MHz, CDCl₃, TMS): 1.55-1.73 (2 H, m, CH₂), 1.74-1.99 (2 H, m, CFHCH₂), 2.48 (3 H, s, CH₃), 2.99 (1 H, m, NCH₂), 3.04–3.17 $(2 \text{ H, m, NCH}_2 \text{ and NC}H_2\text{CHF}), 3.35 (1 \text{ H, ddd}, {}^3J_{\text{H,F}} = 20.2 \text{ Hz},$ $^{2}J_{H,H} = 12.0 \text{ Hz}, ^{3}J_{H,H} = 3.3 \text{ Hz}, \text{NC}H_{2}\text{CFH}), 4.68 (1 \text{ H}, \text{dtt}, ^{2}J_{H,F} =$ 47.2, ${}^{3}J_{H,H} = 6.8$, ${}^{3}J_{H,H} = 3.3$ Hz, CHF), 7.39 (1 H, d, ${}^{3}J_{H,H} = 8.0$ Hz, Ph-CH), 7.61 (1 H, dd, ${}^{3}J_{H,H} = 8.0$, ${}^{4}J_{H,H} = 1.8$ Hz, Ph-CH), 7.94 (1H, d, ${}^{4}J_{H,H}$ = 1.8 Hz, Ph-CH); δ_{C} (101 MHz, CDCl₃) 21.0 (d, ${}^{4}J_{C,F} = 6.3 \text{ Hz}, \text{CH}_{2}\text{CH}_{2}\text{CH}_{2}, 23.1 \text{ (CH}_{3}), 29.2 \text{ (d, } {}^{3}J_{C,F} = 20.1 \text{ Hz},$ CFHCH₂), 45.7 (NCH₂), 49.5 (d, ${}^{2}J_{CF} = 26.5$ Hz, NCH₂CFH), 85.8 (d, ${}^{1}J_{C,F} = 176.8$ Hz, CFH), 125.4 (Ph-C), 126.3 (Ph-CH), 131.1 (Ph-CH), 131.2 (Ph-CH), 135.6 (Ph-C), 143.6 (Ph-C); $\delta_{\rm F}$ (282 MHz, CDCl₃, CFCl₃): -183.5 (1 F, m). MS (EI-GC-inlet): m/z (%) 337/335 (20/21) [M⁺], 337/335 (2/3) [M⁺-HF], 235/233 (10/10) [C₇H₆BrO₂S⁺], 171/169 (12/11) [C₇H₆Br⁺], 102 (100) $[C_5H_9NF^+]$, 90 (22) $[C_7H_6^+]$, 89 (23) $[C_7H_5^+]$, 55 (21) $[C_4H_6^+]$. Exact mass (ESI): [M+Na⁺] calcd for C₁₂H₁₅BrFNO₂SNa⁺: 359.9863/357.9883; found: 359.9871/357.9891.

(2S)-2-Fluoromethyl-N-tosylpyrrolidine (12). Identified by comparison of ¹⁹F NMR data obtained from the reaction mixture of experiment entry 3, Table 2 with known ones; 32,33 $\delta_{\rm F}$ (282 MHz, CDCl₃, CFCl₃): -225.9 (1 F, td, ${}^{2}J_{H,F} = 47.1$ Hz, ${}^{3}J_{H,F} = 16.7$ Hz).

Synthesis of (3R)-3-Fluoro-N-tosylpiperidine (11a) with FluoleadTM and Olah's reagent

The reaction was carried out in a 20 cm³-TeflonTM screwed vessel. To a solution of N-tosly-L-prolinol (3) in absolute dichloromethane (2 cm³) FluoleadTM (250 mg, 1.0 mmol, 2.00 eq.) was added. The vessel was screwed tightly and the reaction mixture was stirred at 85 °C for 45 min. After cooling down to room temperature Olah's reagent $(0.03 \text{ cm}^3, 0.2 \text{ cm}^3 \text{ g}^{-1} \text{ of } 3, 0.22 \text{ eq.})$ was added *via* a polypropylene/polyethylene syringe. The mixture was stirred for 1 h at room temperature, then at 50 °C for 40 min, cooled down to room temperature, neutralized with aqueous NaHCO₃ and extracted with dichloromethane $(3 \times 20 \text{ cm}^3)$. The combined organic layer was washed with 15% aqueous NaOH solution, dried over anhydrous MgSO4 and the solvent was removed under reduced pressure. After column chromatography (silica gel, pentane/diethylether, 40:1) the product 11a was obtained as a white solid (121 mg, 95%). As a by-product (5%, 19F NMR) (2S)-2-fluoromethyl-N-tosylpyrrolidine (12) was found. The spectroscopic data are matching to those given above.

Synthesis of methyl (2S)-2-(1,3-dioxoisoindolin-2-yl)-3-(phenylthio)propanoate (17)

(2S)-2-(1,3-dioxoisoindolin-2-yl)-3-hydroxypropanoic (14)⁴⁸. According to the literature procedure L-serine (1.105 g, 10.0 mmol), sodium carbonate (1.090 g, 10.0 mmol) and Nethoxycarbonylphthalimide (2.190 g, 10.0 mmol) was reacted in water (8 cm³) and the product was obtained as a white solid (2.115 g, 90%); mp 152 °C (with decomposition); $[\alpha]_{D}^{20}$ -8.77 (c 1.00 in CHCl₃); δ_H (300 MHz, D₂O): 1.17 (1 H, t, $^3J_{H,H}$ = 7.1 Hz, CH₂OH), 4.02 (1 H, m, CH₂OH), 4.11 (1 H, m, CH₂OH), 5.00 (1 H, m, NCH), 7.71–7.84 (4 H, m, Ph-CH); $\delta_{\rm C}$ (75 MHz, D₂O): 54.0 (NCH), 58.8 (CH₂OH), 123.4 (Ph-CH), 130.8 (Ph-C), 135.8 (Ph-CH), 169.6 (NCO), 171.5 (COOH). Exact mass (ESI): $[M+Na^{+}]$ calcd for $C_{11}H_{9}NO_{5}Na^{+}$: 258.0373; found: 258.0360. Exact mass (ESI): $[M-H^-]$ calcd for $C_{11}H_8NO_5^-$: 234.0408; found: 234.0406.

(2S)-2-(1,3-dioxoisoindolin-2-yl)-3-hydroxypropano-Methyl ate (15). (2S)-2-(1,3-dioxoisoindolin-2-yl)-3-hydroxypropanoic acid (14) (2.352 g, 10.0 mmol) was dissolved in methanol (20 cm³). Concentrated sulfuric acid (1.07 cm³, 1.960 g, 2.0 mmol, 0.2 eq.) was added and the reaction mixture was stirred at 40 °C for 2 days. Quenching with ice water (50 cm³), extraction with diethyl ether $(3 \times 50 \text{ cm}^3)$, washing of the combined organic layer with saturated NaHCO₃ and water $(2 \times 50 \text{ cm}^3)$, drying over anhydrous MgSO₄ and removing of the solvent under reduced pressure led to the product 15 as a colorless oil (2.240 g, 90%); $[\alpha]_{D}^{20}$ -10.3 (c 1.19 in CHCl₃); $\delta_{\rm H}$ (300 MHz, CDCl₃, TMS): 1.20 (1 H, t, ${}^{3}J_{\rm H,H}$ = 7.1 Hz, CH₂OH), 3.76 (3 H, s, CH₃), 3.85–3.92 (1 H, m, CH₂OH) 4.19-4.25 (1 H, m, CH_2OH), 5.04 (1 H, dd, $^3J_{HH} = 6.1$ Hz, $^3J_{HH} =$ 5.0 Hz, NCH), 7.75 (2 H, dd, ${}^{3}J_{H,H} = 5.6$ Hz, ${}^{4}J_{H,H} = 3.0$ Hz, Ph-CH), 7.86 (2 H, dd, ${}^{3}J_{H,H} = 5.4$ Hz, ${}^{4}J_{H,H} = 3.1$ Hz, Ph-CH); $\delta_{\rm C}$ (75 MHz, CDCl₃): 52.9 (OCH₃), 54.7 (NCH), 61.0 (CH₂OH), 123.7 (Ph-CH), 131.6 (Ph-C), 134.4 (Ph-CH), 168.0 (NCO), 168.4 (COOCH₃). Exact mass (ESI): [M+H⁺] calcd for C₁₂H₁₁NO₅H⁺: 250.0710; found: 250.0708. Exact mass (ESI): [M+Na⁺] calcd for $C_{12}H_{11}NO_5Na^+$: 272.0529; found: 272.0528. Exact mass (ESI): $[M-H^-]$ calcd for $C_{12}H_{10}NO_5^-$: 248.0564; found: 248.0546.

Methyl (2S)-2-(1,3-dioxoisoindolin-2-yl)-3-(phenylthio)propanoate (17). According to the literature procedure³⁹ to a solution of compound 15 (125 mg, 0.5 mmol) in abs. N,Ndimethylformamide tributylphospine (0.19 cm³, 0.75 mmol, 1.5 eq.) and diphenyldisulfide (164 mg, 0.75 mmol, 1.5 eq.) were added. After stirring for 1 h at room temperature the reaction mixture was diluted with diethyl ether (10 cm³), washed with saturated NaHCO₃ solution, water and brine, dried over MgSO₄ and the solvent was removed under reduced pressure. After column chromatography (neutral alumnia, cyclohexane/ethyl acetate, 20:1) the product **17** was obtained as a colorless oil (118 mg, 69%); $[\alpha]_{20}^{20}$ –0.88 (c 1.03 in CHCl₃); $\delta_{\rm H}$ (300 MHz, CDCl₃, TMS): 3.69–3.92 (2 H, m, PhSC H_2), 3.73 (3 H, s, CH₃), 5.01 (1 H, dd, ${}^3J_{\rm H,H}$ = 11.0 Hz, ${}^3J_{\rm H,H}$ = 4.2 Hz, NCH), 7.06 (1 H, m, Ph-CH), 7.16 (2 H, m, Ph-CH), 7.35 (2 H, m, Ph-CH), 7.72 (2 H, m, Ph-CH), 7.79 (2 H, dd, ${}^3J_{\rm H,H}$ = 5.7 Hz, ${}^4J_{\rm H,H}$ = 3.0 Hz, Ph-CH); $\delta_{\rm C}$ (75 MHz, CDCl₃): 33.5 (PhSC H_2), 52.0 (OCH₃), 52.9 (NCH), 123.4 (Ph-CH), 127.0 (Ph-CH), 128.9 (Ph-CH), 131.3 (Ph-CH), 131.5 (Ph-C), 133.6 (Ph-C), 134.0 (Ph-CH), 167.2 (NCO), 168.4 (COOCH₃). Exact mass (ESI): [M+Na⁺] calcd for C₁₈H₁₅NO₄SNa⁺: 364.0614; found: 364.0616.

Oxidative desulfurization—difluorination of methyl (2S)-2-(1,3-dioxoisoindolin-2-yl)-3-(phenylthio)propanoate (17)

According to above general procedure for the oxidative desulfurization—fluorination, methyl (2*S*)-2-(1,3-dioxoisoindolin-2-yl)-3-(phenylthio)-propanoate (17) (86 mg, 0.25 mmol) in dry dichloromethane (5 cm³) was reacted with Olah's reagent (0.35 cm³, 1.5 mmol, 6 eq.) and DBH (215 mg, 0.75 mmol, 3 eq.) while stirring for 30 min at 0 °C and at room temperature over night. Then the reaction mixture was neutralized with saturated NaHCO₃ solution and extracted with dichloromethane (3 × 10 cm³). The combined organic layer was washed with 1 N HCl, 5% aqueous NaHCO₃, dried over anhydrous MgSO₄ and the solvent was removed under reduced pressure. After column chromatography (neutral alumina, cyclohexane/ethyl acetate, 20:1) a lightly yellow solid was obtained. Recrystallization from pentane—diethyl ether gave colorless crystals of compound 19, which were subjected to X-ray crystallography.

Methyl (2*S*)-2-(1,3-dioxoisoindolin-2-yl)-3,3-diffuoropropionate (18). The formed minor component 18 could only be detected by ESI mass spectrometry and ¹⁹F NMR spectroscopy. δ_F (282 MHz, CDCl₃, CFCl₃): -118.0 (2 F, ddd, ² J_{FF} = 5.5 Hz, ² J_{HF} = 63.0 Hz, ³ J_{HF} = 21.3 Hz). Exact mass (ESI): [M+Na⁺] calcd for C₁₂H₉F₅NO₄Na⁺: 292.0392; found: 292.0380.

Methyl 2,2-dibromo-2-(1,1-difluoro-3-oxoisoindolin-2-yl)acetate (19). (21 mg, 21%); mp 125–126 °C; $\delta_{\rm H}$ (500 MHz, CDCl₃, TMS): 3.96 (3 H, s, CH₃), 7.73 (1 H, m, Ph-CH), 7.78-7.83 (2 H, m, Ph-CH), 7.85–7.89 (1 H, d, ${}^{3}J_{H,H} = 7.6$ Hz, Ph-CH); $\delta_{\rm C}$ (126 MHz, CDCl₃): 51.8 (t, ${}^{3}J_{\rm C,F}$ = 1.2 Hz, Br₂NCCO), 55.4 (CH₃), 120.4 (t, ${}^{1}J_{CF} = 256.8$ Hz, CF₂N), 122.5 (Ph-CH), 124.5 (Ph-CH), 127.6 (t, ${}^{3}J_{C,F} = 2.3$ Hz, Ph-C), 132.9 (Ph-C), 135.2 (Ph-CH), 138.0 (t, ${}^{2}J_{C,F} = 26.1$ Hz, Ph-C), 163.3 $(COOCH_3)$, 163.6 (NCO); δ_F (470 MHz, CDCl₃, CFCl₃): -87.0 (2 F, s). MS (EI-GC-inlet): m/z (%) 401/399/397 (<0.1/<0.1/<0.1) $[M^+]$, 370/368/366 (1/2/1) $[M^+-CH_3O]$, 342/340/338 (4/9/4) $[M^+-C_2H_3O_2]$, 320/318 (100/98) $[M^+-Br]$, 235/233 (66/68), 180 (38) $[C_{11}H_7Br_2F_2NO_3^+]$, 154 (72), 152 (47) $[C_8H_4F_2N^+]$, 126 (89) $[C_4H_7F_2^+]$, 125 (66). Exact mass (ESI): $[M+Na^+]$ calcd for $C_{11}H_7Br_2F_2NO_3Na^+$: 423.8612/421.8633/419.8653; found: 423.8615/421.8635/419.8659.

Crystallographic data

Crystal structure analysis of compound **19**: $C_{11}H_7Br_2F_2NO_3$, M = 399.00, colorless crystal $0.40 \times 0.10 \times 0.01$ mm, a = 13.5963(9), b = 7.1431(5), c = 13.1021(9) Å, $\beta = 94.214(3)^\circ$, V = 1269.03(15) Å³, $\rho_c = 2.088$ g cm⁻³, $\mu = 8.408$ mm⁻¹, empirical absorption

Data set was collected with a Nonius KappaCCD diffractometer. Programs used: data collection COLLECT (Nonius B.V., 1998), data reduction Denzo-SMN,⁴⁹ absorption correction Denzo,⁵⁰ structure solution SHELXS-97,⁵¹ structure refinement SHELXL-97,⁵² graphics SCHAKAL (E. Keller, Univ. Freiburg, 1997).

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Notes and references

- (a) R. W. Fuller and B. B. Molly, The Effect of Aliphatic Fluorine on Amine Drugs, in Biochemistry Involving Carbon-Fluorine Bonds, ed. R. Fuller, ACS Symposium Series, Vol. 28, American Chemical Society, Washington, 1976, pp. 77–98; (b) M. B. van Niel, M. S. Beer, H. B. Broughton, S. K. F. Cheng, S. S. Goodacre, A. Heald, K. L. Locker, A. M. MacLeod, D. Morrison, C. R. Moyes, D O'Connor, A. Pike, M. Rowley, M. G. N. Russel, B. Sohal, J. A. Stanton, S Thomas, H. Verrier, A. P. Watt and J. L. Castro, J. Med. Chem., 1999, 42, 2087–2104; (c) A. L. Upthagrove and W. L. Nelson, Drug Metabol. Disp., 2001, 29, 1389–1395; (d) M. Morgenthaler, E. Schweizer, A. Hoffmann-Röder, F. Bonini, R. E. Martin, G. Jaeschke, B. Wagner, H. Fischer, S. Bendels, D. Zimmerli, J. Schneider, F. Diederich, M. Kansy and K. Müller, ChemMedChem, 2007, 2, 1100–1115.
- Recent reviews: (a) H.-J. Böhm, D. Banner, S. Bendels, M. Kansy, B. Kuhn, K. Müller, U. Obst-Sander and M. Stahl, ChemBioChem, 2004, 5, 637–643; (b) C. Isanbor and D. O'Hagan, J. Fluorine Chem., 2006, 127, 303–319; (c) J. P. Bégué and D. Bonnet-Delpon, J. Fluorine Chem., 2006, 127, 992–1012; (d) K. L. Kirk, J. Fluorine Chem., 2006, 127, 1013–1029; (e) K. Müller, C. Faeh and F. Diederich, Science, 2007, 317, 1881–1886; (f) W. K. Hagmann, J. Med. Chem., 2008, 51, 4359–4369.
- 3 Recent books: (a) J.-P. Bégué and D. Bonnet-Delpon, Bioorganic and Medicinal Chemistry of Fluorine, John Wiley & Sons, 2008; (b) Fluorine and Health. Molecular Imaging, Biomedical Materials and Pharmaceuticals, ed. A. Tressaud and G. Haufe, Elsevier, Amsterdam, 2008; (c) Fluorine in Medicinal Chemistry and Chemical Biology, ed. I. Ojima, Wiley-Blackwell, Chichester, 2009.
- 4 P. Bey, J. Fozard, J. M. Lacoste, I. A. McDonald, M. Zreika and M. G. Palfreyman, *J. Med. Chem.*, 1984, **27**, 9–10.
- 5 (a) S. Yoshida, O. G. J. Meyer, T. C. Rosen, S. Ye, M. J. Sloan, G. Haufe and K. L. Kirk, J. Med. Chem., 2004, 47, 1796–1806; (b) S. Yoshida, T. C. Rosen, O. G. J. Meyer, S. Ye, M. J. Sloan, G. Haufe and K. L. Kirk, Bioorg. Med. Chem., 2004, 12, 2645–2652; (c) T. C. Rosen, S. Yoshida, K. L. Kirk and G. Haufe, J. Med. Chem., 2004, 47, 5860–5871; (d) S. Ye, S. Yoshida, R. Fröhlich, G. Haufe and K. L. Kirk, Bioorg. Med. Chem., 2005, 13, 2489–2499; (e) S. Hruschka, T. C. Rosen, S. Yoshida, K. L. Kirk, R. Fröhlich, B. Wibbeling and G. Haufe, Bioorg. Med. Chem., 2008, 16, 7148–7166; (f) S. Hruschka, S. Yoshida, K. L. Kirk and G.

- Haufe, J. Fluorine Chem., 2008, 129, 875-880; (g) I. Hyla-Kryspin, S. Grimme, S. Hruschka and G. Haufe, Org. Biomol. Chem., 2008, 6,
- 6 L. A. Sorbera, M. del Fresno and X. Rabasseda, Drugs Future, 2003, 28, 217-223.
- 7 (a) Fluorine-containing Amino Acids. Synthesis and Properties. V. P. Kukhar and V. A. Soloshonok, ed., John Wiley & Sons, Chichester, 1995; (b) S. Kröger and G. Haufe, Amino Acids, 1997, 12, 363-372; (c) V. Tolman, Amino Acids, 1996, 11, 15-36; (d) A. Sutherland and C. L. Willis, Nat. Prod. Rep., 2000, 17, 621-631; (e) X.-L. Qui, W.-D. Meng and F.-L. Qing, Tetrahedron, 2004, 60, 6711-6745.
- 8 N. Fokina, A. Kornilov, I. Kulik and V. Kukhar, J. Fluorine Chem., 2001, 111, 69-76.
- 9 K. Uoto, S. Ohsuki, H. Takenoshita, T. Ishiyama, S. Iimura, Y. Hirota, I. Mitsui, T. Hirofumi and T. Soga, Chem. Pharm. Bull., 1997, 45, 1793-1804.
- 10 K. Nakayama, H. C. Kawato, H. Inagaki, R. Nakajima, A. Kitamura, K. Someya and T. Ohta, Org. Lett., 2000, 2, 977-980.
- 11 P.G. Cozzi, Angew. Chem., Int. Ed., 2007, 46, 2568–2571; Angew. Chem., 2007, 119, 2620-2623.
- 12 R. Ocampo and W. R. Dolbier Jr., Tetrahedron, 2004, 60, 9325-9374.
- 13 S. Marcotte, X. Pannecoucke, C. Feasson and J.-C. Quirion, J. Org. Chem., 1999, 64, 8461-8464.
- 14 A. Sorochinsky, N. Voloshin, A. Markovsky, M. Belik, N. Yasuda, H. Uekusa, T. Ono, D. O. Berbasov and V. A. Soloshonok, J. Org. Chem., 2003, 68, 7448-7454.
- 15 V. A. Soloshonok, H. Ohkura, A. Sorochinsky, N. Voloshin, A. Markovsky, M. Belik and T. Yamazaki, Tetrahedron Lett., 2002, 43, 5445-5448.
- 16 W. Metcalf, P. Bey, C. Danzin, M. J. Jung, P. Casara and J. P. Vevert, J. Am. Chem. Soc., 1978, 100, 2551-2699.
- 17 Y. Ding, J. Wang, K. A. Abbound, Y. Xu, W. R. Dolbier Jr. and N. G. J. Richards, J. Org. Chem., 2001, 66, 6381-6388.
- 18 N. Fokina, A. Kornilov, I. Kulik and V. Kukhar, Synthesis, 2002, 2589-
- 19 T. Ohba, E. Ikeda and H. Takei, Bioorg. Med. Chem. Lett., 1996, 6, 1875-1880.
- 20 J. Kollonitsch, S. Marburg and L. M. Perkins, J. Org. Chem., 1976, 41, 3107-3111.
- 21 J. Kollonitsch, L. Barash and G. A. Douldoras, J. Am. Chem. Soc., 1970, 92, 7494-7495.
- 22 J. J. M. Hagemann, M. J. Wanner, G. J. Koomen and U. K. Pandit, J. Med. Chem., 1977, 20, 1677-1679.
- 23 F. N. Shirota, H. T. Nagasawa and J. A. Elberling, J. Med. Chem., 1977, 20, 1623-1627.
- 24 H. Hoshi, S. Aburaki, S. Imura, T. Yamasaki, T. Naito and H. Kawaguchi, J. Antibiotics, 1990, 43, 858-872
- 25 A. K. Podichetty, A. Faust, K. Kopka, S. Wagner, O. Schober, M. Schäfers and G. Haufe, Bioorg. Med. Chem., 2009, 17, 2680-2688.
- 26 A. K. Podichetty, S. Wagner, A. Faust, M. Schäfers, O. Schober, K. Kopka and G. Haufe, Future Med. Chem., 2009, 1, 969-989.
- 27 (a) K. C. Nicolaou, R. E. Dolle, D. P. Papahatjis and J. L. Randall, J. Am. Chem. Soc., 1984, 106, 4189-4192; (b) S. C. Sondej and J. A. Katzenellenbogen, J. Org. Chem., 1986, 51, 3508-3513; (c) M. Shimizu, T. Maeda and T. Fujisawa, J. Fluorine Chem., 1995, 71, 9-12; (d) S. Caddick, L Gazzard, W. B. Motherwell and J. A. Wilkinson, Tetrahedron, 1996, 52, 149-156; (e) C. York, G. K. S. Prakash and G. A. Olah, *Tetrahedron*, 1996, **52**, 9–14; (f) S. Furuta, M. Kuroboshi and

- T. Hiyama, Bull. Chem. Soc. Jpn., 1998, 71, 2687–2694; (g) A. Hagooly and S. Rozen, J. Org. Chem., 2004, 69, 7241-7245.
- 28 (a) V. Hugenberg and G. Haufe, Synlett, 2009, 106-108; (b) V. Hugenberg and G. Haufe, J. Fluorine Chem., 2010, 131, 942-950; (c) V. Hugenberg, S. Wagner, K. Kopka, O. Schober, M. Schäfers and G. Haufe, J. Org. Chem., 2010, 75, 6086-6095.
- 29 G. V. Shustov, S. N. Denisenko, I. I. Chervin and R. G. Kostyanovskii, Bull. Acad. Sci. USSR, Div. Chem. Sci. (Engl. Transl.), 1988, 37, 1422 1427; D. S. Radchenko, P. K. Mykhailiuk, A. V. Bezdudny and I. V. Komarov, Synlett, 2009, 1827-1829.
- 30 (a) M. Kuroboshi, K. Kanie and T. Hiyama, Adv. Synth. Catal., 2001, 343, 235-244; (b) M. Shimizu and T. Hiyama, Angew. Chem., 2005, 117, 218–234; Angew. Chem., Int. Ed., 2005, 44, 214–231; (c) S. Ayuba, N. Yoneda, T. Fukuhara and S. Hara, Bull. Chem. Soc. Jpn., 2002, 75, 1597-1603; (d) S. Ayuba, T. Fukuhara and S. Hara, Org. Lett., 2003, **5**, 2873–2874; (e) S. Ayuba, C. Hiramatsu, T. Fukuhara and S. Hara, Tetrahedron, 2004, 60, 11445-11451.
- 31 U.S. Pat. Appl. Publ. US 2008039660 A1 20080214, 2008.
- 32 C. Ye and J. M. Shreeve, J. Fluorine Chem., 2004, 125, 1869–1872
- 33 (a) I. Déchamps, D. Gomez Pardo and J. Cossy, Eur. J. Org. Chem., 2007, 4224-4234; (b) I. Déchamps, D. Gomez Pardo and J. Cossy, Synlett, 2007, 263-267.
- 34 I. I. Gerus, A. A. Kolomeitsev, M. I. Kolycheva and V. P. Kukhar, J. Fluorine Chem., 2000, 105, 31-33.
- 35 T. Tsushima, K. Kawada, S. Iahihara, N. Uchida, O. Shiratori, J. Higaki and M. Hirata, Tetrahedron, 1988, 44, 5375-5387.
- 36 T. Tsushima and K. Kawada, *Tetrahedron Lett.*, 1985, **26**, 2445–2448.
- 37 H. d'Orchymont, Synthesis, 1993, 961-963.
- 38 M.-Y. Chang, S.-T. Chen and N.-C. Chang, Synth. Commun., 2003, 33, 1375-1382.
- 39 I. Nakagawa and T. Hata, Tetrahedron Lett., 1975, 16, 1409-1412.
- 40 W. Dmowski, Methods of Organic Chemistry (Houben-Weyl), Vol. E10a, ed. B. Baasner, H. Hagemann and J. C. Tatlow, Thieme Stuttgart, 2000, 336-344.
- 41 M. H. Rock, Methods of Organic Chemistry (Houben-Weyl), Vol. E10a, ed. B. Baasner, H. Hagemann and J. C. Tatlow, Thieme, Stuttgart, 2000,
- 42 V. Hugenberg, PhD Thesis, University of Münster, 2009.
- 43 C. Z. Dong, A. Ahamada-Himidi, S. Plocki, D. Aoun, M. Touaibia, N. M. B. Habich, C. Redeuilh, J. E. Ombetta, J. J. Godfrold, F. Massicot and F. Heymans, Bioorg. Med. Chem., 2005, 13, 1989-2007.
- 44 W. Zhang, Y. Qin, S. Zhang and M. Luo, ARKIVOC 2005, Part (xiv): General Papers, 39-48.
- 45 E. D. Nicolaides, F. J. Tinney, J. S. Kaltenbronn, J. T. Repine, D. A. DeJohn, E. A. Lunney and W. H. Roark, J. Med. Chem., 1986, 29, 959-971.
- 46 J. A. Katzenellenbogen, J. Fluorine Chem., 2001, 109, 49-54.
- 47 D. C. Craig, G. L. Edwards and C. A. Muldoon, Synlett, 1997, 1441-
- 48 G. H. Nefken, G. I. Tesser and R. J. F. Nivard, Recl. Trav. Chim. Pays-Bas., 1960, 79, 688-698.
- 49 Z. Otwinowski and W. Minor, Methods Enzymol., 1997, 276, 307-326.
- 50 Z. Otwinowski, D. Borek, W. Majewski and W. Minor, Acta Crystallogr., Sect. A: Found. Crystallogr., 2003, 59, 228-234.
- 51 G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Crystallogr., 1990, 46, 467-473.
- 52 G. M. Sheldrick, Acta Cryst., 2008, A64, 112-122.